

International concern is growing regarding antimicrobial drug resistance in *Shigella* infections associated with India. Fluoroquinolone resistance emerged in *S. dysenteriae* in 2002, in *S. flexneri* in 2004, and in *S. sonnei* in 2007 (5). Studies from Japan have also reported an association between travel to India and infection with an *S. sonnei* clonal group that was multidrug resistant, including resistance to nalidixic acid (6). Furthermore, ciprofloxacin-resistant *S. sonnei* isolates from foodborne outbreaks in India in 2009 and 2010 (7) had *Xba*I- PFGE types and resistance profiles visually indistinguishable from those reported in our study. A study of *S. sonnei* isolates in Bhutan showed that this clonal group was also common there (8). Furthermore, a 2010 outbreak of ciprofloxacin-resistant *S. sonnei* in Canada associated with men who have sex with men showed *Xba*I- and *Bln*I-PFGE patterns that appear similar to the patterns for isolates in this study (9).

Antimicrobial drug resistance is a major global problem that is likely to be exacerbated in places with poor sanitation and intensive use of antimicrobial drugs in humans and animals. These factors have contributed to increased ciprofloxacin resistance in *Salmonella enterica* serovars Typhi and Paratyphi A (10).

A review of published literature and informal communication indicates that our observation of ciprofloxacin resistance in *S. sonnei* infections associated with travel to India is part of a general global trend. This increasing resistance suggests that ciprofloxacin may no longer be suitable for empiric therapy for *S. sonnei* infection, particularly for patients with a history of travel to the subcontinent of India.

## References

- Holt KE, Baker S, Weill FX, Holmes E, Kitchen A, Yu J, et al. *Shigella sonnei* genome sequencing and phylogenetic analysis indicate recent global dissemination from Europe. *Nat Genet*. 2012;44:1056–9. <http://dx.doi.org/10.1038/ng.2369>
- DeLappe N, O'Connor J, Morris D, Cormican M. Molecular detection of *Shigella* species impacts on apparent epidemiology and reference laboratory workload. In: Final Program of the 24th European Congress of Clinical Microbiology and Infectious Diseases; Barcelona, Spain; 2014 May 10–13; ePoster 091. Basel (Switzerland): European Society of Clinical Microbiology and Infectious Diseases; 2014.
- Ribot EM, Fair MA, Gautom R, Cameron DN, Hunter SB, Swaminathan B. Standardization of pulsed-field gel electrophoresis protocols for the subtyping of *Escherichia coli* O157:H7, *Salmonella* and *Shigella* for PulseNet. *Foodborne Pathog Dis*. 2006;3:59–67. <http://dx.doi.org/10.1089/fpd.2006.3.59>
- World Health Organization. Guidelines for the control of shigellosis, including epidemics of *Shigella dysenteriae* type 1. 2005 [cited 2015 Mar 4]. <http://whqlibdoc.who.int/publications/2005/9241592330.pdf>
- Nandy S, Mitra U, Rajendran K, Dutta P, Dutta S. Subtype prevalence, plasmid profiles and growing fluoroquinolone resistance in *Shigella* from Kolkata, India (2001–2007): a hospital-based study. *Trop Med Int Health*. 2010;15:1499–507. <http://dx.doi.org/10.1111/j.1365-3156.2010.02656.x>
- Izumiya H, Tada Y, Ito K, Morita-Ishihara T, Ohnishi M, Terajima J, et al. Characterization of *Shigella sonnei* isolates from travel-associated cases in Japan. *J Med Microbiol*. 2009;58:1486–91. <http://dx.doi.org/10.1099/jmm.0.011809-0>
- Nandy S, Dutta S, Ghosh S, Ganai A, Jyothi R, Ramani Bai JT, et al. Foodborne-associated *Shigella sonnei*, India, 2009 and 2010. *Emerg Infect Dis*. 2011;17:2072–4. <http://dx.doi.org/10.3201/eid1711.110403>
- Ruekit S, Wangchuk S, Dorji T, Tshering KP, Pootong P, Nobthai P, et al. Molecular characterization and PCR-based replicon typing of multidrug resistant *Shigella sonnei* isolates from an outbreak in Thimphu, Bhutan. *BMC Res Notes*. 2014;7:95.
- Gaudreau C, Ratnayake R, Pilon PA, Gagnon S, Roger M, Lévesque S. Ciprofloxacin-resistant *Shigella sonnei* among men who have sex with men, Canada, 2010. *Emerg Infect Dis*. 2011;17:1747–50. <http://dx.doi.org/10.3201/eid1709.102034>
- Dutta S, Das S, Mitra U, Jain P, Roy I, Ganguly S, et al. Antimicrobial resistance, virulence profiles and molecular subtypes of *Salmonella enterica* serovars Typhi and Paratyphi A blood isolates from Kolkata, India during 2009–2013. *PLoS ONE*. 2014;9:e101347. <http://dx.doi.org/10.1371/journal.pone.0101347>

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## Fatal *Balamuthia mandrillaris* Meningoencephalitis in the Netherlands after Travel to The Gambia

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**To the Editor:** *Balamuthia mandrillaris* is a free-living amoeba that has a worldwide distribution in soil and was first reported in 1990 (1). Approximately 200 *B. mandrillaris* meningoencephalitis cases have been described, mostly from warm climate areas in South America. Its prevalence in the United States is estimated to be 1 case/year (2). However, *B. mandrillaris* meningoencephalitis

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has not been reported in Africa, and only 4 cases have been reported in Europe (3–6). Transmission occurs through the respiratory tract or the skin or by organ transplant, and the incubation period varies from weeks to months after primary infection (7). After an indolent, subacute phase with aspecific symptoms, the amebae invade the central nervous system, and illness rapidly progresses, leading almost invariably to death (7). Because *B. mandrillaris* is difficult to detect in soil, its specific geographic distribution around the world is unknown and is estimated on the basis of where illnesses have been reported (7). This report addresses fatal *B. mandrillaris* meningoencephalitis in a woman from the Netherlands who had visited The Gambia.

In December 2013, a previously healthy 61-year-old white woman in the Netherlands sought care for fever, headaches, and muscle pains she had experienced for 1 week. That year, she had traveled 4 times to The Gambia, the last visit being 1 month before her hospitalization (online Technical Appendix Table 1, <http://wwwnc.cdc.gov/EID/article/21/5/14-1325-Techapp1.pdf>). After she returned from her visit in September 2013, fatigue, diarrhea, fever, and pustular skin lesions on her back and lower extremities developed. A wound swab culture showed *Staphylococcus aureus*, for which she was treated successfully with oral clarithromycin and topical fucidin ointment.

On admission in December, her physical and neurologic examination results were unremarkable. Malaria was excluded; because of persisting headaches, a cerebral computed tomography scan without contrast was performed but showed no abnormalities. In the following days, high fevers, altered mental status, and nuchal rigidity without focal neurologic deficits developed. Cerebrospinal fluid (CSF) examination showed mononuclear pleocytosis, highly elevated protein levels, and low glucose levels (online Technical Appendix Table 2). Serial cerebral computed tomography and magnetic resonance imaging scans showed development of an asymmetric hydrocephalus and diffuse leptomeningeal and subependymal contrast enhancement,

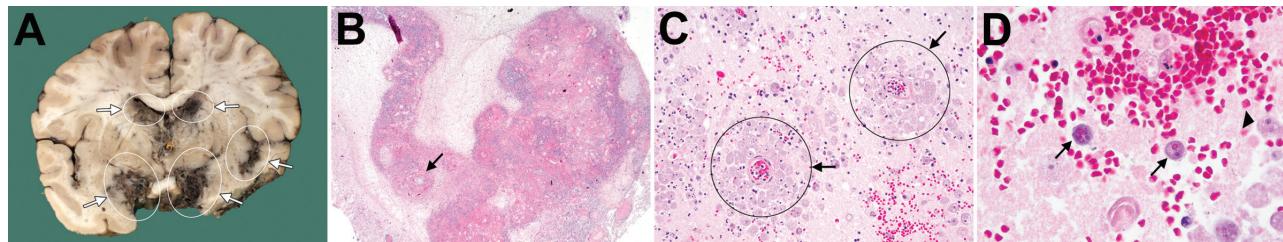
especially around the brainstem, without signs of intracerebral mass lesions (online Technical Appendix Figure).

Presumed diagnosis was tuberculous meningitis, and she was treated with tuberculostatic drugs (isoniazid, rifampin, pyrazinamide, and ethambutol) combined with intravenous acyclovir, ceftriaxone, and co-trimoxazole for other infectious causes of meningoencephalitis. Despite external lumbar and ventricular (both lateral ventricles and fourth ventricle) CSF drainage, her neurologic condition deteriorated. Multiple cranial nerve palsies developed, and she became comatose and died 11 days after admission.

Informed consent for postmortem examination was obtained, and macroscopic pathologic examination showed uncal and cerebellar herniation caused by increased intracranial pressure. Microscopic brain tissue examination showed signs of acute granulomatous inflammation, multiple hemorrhagic infarctions, and angiitis in the presence of numerous amebic trophozoites and cysts (Figure), which showed granulomatous hemorrhagic necrotic amebic meningoencephalitis. Real-time PCR and subsequent sequencing on brain biopsy and CSF specimens showed *B. mandrillaris* to be the causative ameba (8,9).

The infection could have been acquired in The Gambia or the Netherlands because the patient had intensive soil contact in The Gambia, where she frequently cultivated land, and in the Netherlands, where she worked in glass horticulture. She may have been infected through the skin after contact with contaminated soil, but her skin lesions were atypical for *B. mandrillaris*, and postmortem examinations failed to identify *B. mandrillaris* except in the central nervous system.

The lack of reported *B. mandrillaris* cases from Africa might indicate a low number of postmortem examinations and little access to advanced diagnostics, rather than a low environmental prevalence of *B. mandrillaris*. The few reported cases in Europe might be related to lack of awareness and to clinical signs and symptoms that mimic tuberculous meningitis: a lymphocytic pleocytosis with an elevated protein level and a low glucose level in CSF, together with a hydrocephalus and subependymal and



**Figure.** Postmortem pathologic findings for woman in the Netherlands who died of *Balamuthia mandrillaris* meningoencephalitis after returning from travel to The Gambia. A) Macroscopic coronal central section scan showing hemorrhagic necrotizing lesions of the subependymal, meningeal, and parenchymal areas of the parietotemporal lobes (circles and arrows). B) Low-power microscopic scan showing hemorrhagic necrotizing angiitis of the meningeal vessels (arrow) (original magnification  $\times 25$ ). C) Medium-power microscopic scan (original magnification  $\times 200$ ) showing perivascular trophozoite cuffing (arrows) and granulomatous inflammation. D) High-power microscopic scan (original magnification  $\times 630$ ) showing encysted amebae (arrows) and free trophozoites (arrowhead). Hematoxylin and eosin stains.

leptomeningeal contrast enhancement on magnetic resonance imaging (10). Also, *B. mandrillaris* meningoencephalitis imaging findings are often nonspecific, including cerebral edema, hydrocephalus, multiple space-occupying and ring-enhancing lesions, leptomeningeal enhancement, or formation of mycotic aneurysms (2). Furthermore, amebic trophozoites are seldom detected in CSF by microscopy (2,3). Consequently, *B. mandrillaris* meningoencephalitis could be underdiagnosed, especially where this infection has no or only sporadic reports.

*B. mandrillaris* should be considered in refractory or unexplained cases of meningoencephalitis, even outside the Americas and in immunocompetent patients. Detecting *B. mandrillaris* by PCR in CSF seems most likely to enable early diagnosis and timely treatment. However, appropriate therapy is not well defined; success has been sparsely reported with the simultaneous use of azoles, flucytosine, pentamidine, sulfazidine, macrolide antimicrobial drugs, phenothiazines, and miltefosine (2,7,10).

## References

1. Visvesvara GS, Martinez AJ, Schuster FL, Leitch GJ, Wallace SV, Sawyer TK, et al. Leptomyxid ameba, a new agent of amebic meningoencephalitis in humans and animals. *J Clin Microbiol.* 1990;28:2750–6.
2. Diaz JH. The public health threat from *Balamuthia mandrillaris* in the southern United States. *J La State Med Soc.* 2011;163:197–204.
3. Jayasekera S, Sissons J, Tucker J, Rogers C, Nolder D, Warhurst D, et al. Post-mortem culture of *Balamuthia mandrillaris* from the brain and cerebrospinal fluid of a case of granulomatous amoebic meningoencephalitis, using human brain microvascular endothelial cells. *J Med Microbiol.* 2004;53:1007–12. <http://dx.doi.org/10.1099/jmm.0.45721-0>
4. Kodet R, Nohynkova E, Tichy M, Soukup J, Visvesvara GS. Amebic encephalitis caused by *Balamuthia mandrillaris* in a Czech child: description of the first case from Europe. *Pathol Res Pract.* 1998;194:423–9. [http://dx.doi.org/10.1016/S0344-0338\(98\)80033-2](http://dx.doi.org/10.1016/S0344-0338(98)80033-2)
5. Tavares M, Correia da Costa JM, Carpenter SS, Santos LA, Afonso C, Aguiar A, et al. Diagnosis of first case of *Balamuthia* amoebic encephalitis in Portugal by immunofluorescence and PCR. *J Clin Microbiol.* 2006;44:2660–3. <http://dx.doi.org/10.1128/JCM.00479-06>
6. White JM, Barker RD, Salisbury JR, Fife AJ, Lucas SB, Warhurst DC, et al. Granulomatous amoebic encephalitis. *Lancet.* 2004;364:220. [http://dx.doi.org/10.1016/S0140-6736\(04\)16640-3](http://dx.doi.org/10.1016/S0140-6736(04)16640-3)
7. Matin A, Siddiqui R, Jayasekera S, Khan NA. Increasing importance of *Balamuthia mandrillaris*. *Clin Microbiol Rev.* 2008;21:435–48. <http://dx.doi.org/10.1128/CMR.00056-07>
8. Qvarnstrom Y, Visvesvara GS, Sriram R, da Silva AJ. Multiplex real-time PCR assay for simultaneous detection of *Acanthamoeba* spp., *Balamuthia mandrillaris*, and *Naegleria fowleri*. *J Clin Microbiol.* 2006;44:3589–95. <http://dx.doi.org/10.1128/JCM.00875-06>
9. Kiderlen AF, Radam E, Lewin A. Detection of *Balamuthia mandrillaris* DNA by real-time PCR targeting the RNase P gene. *BMC Microbiol.* 2008;8:210. <http://dx.doi.org/10.1186/1471-2180-8-210>
10. Visvesvara GS, Moura H, Schuster FL. Pathogenic and opportunistic free-living amoebae: *Acanthamoeba* spp., *Balamuthia mandrillaris*, *Naegleria fowleri*, and *Sappinia diploidea*. *FEMS Immunol Med Microbiol.* 2007;50:1–26. <http://dx.doi.org/10.1111/j.1574-695X.2007.00232.x>

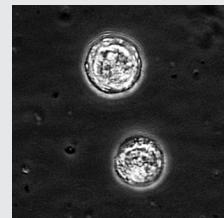
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# etymologia

## *Balamuthia mandrillaris* [bal"ə-moo'the-ə man"dril-a'ris]

A free-living ameba naturally found in the environment, *Balamuthia mandrillaris* can cause a serious infection of the brain, other organs (skin, liver, kidneys), and rarely, spinal cord. Originally isolated from the brain of a mandrill that died of meningoencephalitis at the San Diego Zoo, *Balamuthia mandrillaris* is named for the late professor

William Balamuth of the University of California at Berkeley, for his contributions to the study of amebae. More recently, *B. mandrillaris* has been shown to be transmissible through organ transplantation.



## Sources

1. Centers for Disease Control and Prevention. *Balamuthia mandrillaris*—granulomatous amebic encephalitis (GAE) [cited 2015 Feb 10]. <http://www.cdc.gov/parasites/balamuthia/>
2. Centers for Disease Control and Prevention. *Balamuthia mandrillaris* transmitted through organ transplantation—Mississippi, 2009. *MMWR Morb Mortal Wkly Rep.* 2010;59:1165–70.
3. Schuster FL. In memoriam: William Balamuth (1914–1981). *J Protozool.* 1982;29:1–2. <http://dx.doi.org/10.1111/j.1550-7408.1982.tb02872.x>
4. Visvesvara GS, Schuster FL, Martinez AJ. *Balamuthia mandrillaris*, n. g., n. sp., agent of amebic meningoencephalitis in humans and other animals. *J Eukaryot Microbiol.* 1993;40:504–14. <http://dx.doi.org/10.1111/j.1550-7408.1993.tb04943.x>

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